



Accredited Veterinarian Newsletter

JAN/FEB 2010

NVAP Changes

By Dr. Rochelle Woods, USDA



The U.S. Department of Agriculture has updated the veterinary accreditation program. The National Veterinary Accreditation Program is a voluntary program that is based on a partnership the Animal and Plant Health Inspection Service has with State Animal Health officials and accredited veterinarians. Through the accreditation program, the USDA tries to meet the needs for our national official veterinary infrastructure in the complex and fast paced world of animal health, business, trade and travel. The changes to the program as of December 2009, include:

- Veterinarians who are already accredited must elect to participate in the updated accreditation program by August 2010 or their accreditation will expire.
- Veterinarians who are already accredited will have to choose an accreditation category (Category I or Category II).
- Accredited veterinarians will be issued a new 6 digit national accreditation number that will replace all accreditation numbers previously assigned by state or federal officials. This number will have no correlation to the state license number.
- Accredited veterinarians will have to renew their accreditation every 3 years.
- Accredited veterinarians will have to complete APHIS approved supplemental training in order to renew their accreditation. 3 units of supplemental training will be needed to renew Category I accreditation. 6 units of supplemental training will be needed to renew Category II accreditation.
- For veterinarians who have never been accredited, beginning in July 1, 2011, applicants for accreditation will have to complete specific APHIS approved initial accreditation training through a US college of veterinary medicine or through the Association for American Veterinary Medical Colleges. Beginning July 1, 2011, a certificate of completion of this training will be required for an applicant to be admitted to the Core Orientation.
- Requirements for initial accreditation are:
 - Complete Core Orientation
 - Graduate with a doctor of veterinary medicine degree or the equivalent
 - Be licensed or legally able to practice in the state in which you want to be accredited
 - Submit the National Veterinary Accreditation Program Application Form ([VS 1-36A](#))
 - Complete State-specific training if required
 - Obtain written approval to perform accredited duties from the Animal and Plant Health Inspection Service
- The National Veterinary Accreditation Program Application Form ([VS 1-36A](#)) is established as a multiple use form for accredited veterinarians to update their contact information, request a change in accreditation category, etc. We will continue to provide more detailed information in upcoming Accredited Vet Newsletters. In the meantime, remember to send in your [VS 1-36A](#) form to elect participation in the accreditation program.
- Be sure to check out the NVAP website below for full details of the program, required online courses, forms, FAQs, etc.



<http://www.aphis.usda.gov/nvap/>

IN THIS ISSUE: A New Beginning, CEM Surveillance Study, New Johne's Tests, Anaplasmosis Policy Change, Going Green, 2010 Fair Information, Cattle Ear Tags and Canadian Importation, CEM Biosecurity, Tuberculin Shortage, Import/Export Website, Rabid Dairy Calf with Vesicles, Donkey Deaths

A New Beginning...

By DAI Staff



After 30+ years of state service, with 20 of those years as state veterinarian, Dr. John Huntley is leaving the Division of Animal Industry. Dr. Huntley will begin a new chapter in his veterinary career with the USDA-APHIS-VS. He will be the AVIC (Area Veterinarian in Charge) of Washington State, Hawaii and Alaska. Dr. Huntley leaves New York after many years of working for positive change for animal agriculture in the state. He has tirelessly worked to create strong bonds between producers, veterinarians and regulatory officials. He also has successfully orchestrated the eradication of several diseases from the state. While serving as state veterinarian, Dr. Huntley was deployed to Iraq. His work overseas reminds us that veterinary medicine encompasses many aspects and has no borders. For those of us at the Division of Animal Industry who had the privilege to work with him, he has been a sound leader. He is inspiring, compassionate, honest and most importantly, supportive of the goals of the division. Many of us seem him equally as a friend as well as director. We wish him all the best as he heads westward to start his career with the federal government. He will be missed!

Reminder to NYSCHAP certified and herd veterinarians:

Submit your vouchers promptly! To qualify for payment, vouchers must be received by the Department **no later than 90 days** following the date of the herd visit.



USDA-APHIS-VS CEM Stallion Surveillance Study

USDA-APHIS-VS will begin **voluntary** CEM surveillance testing of 4,000 stallions starting around February 1, 2010. Stallions will be cultured for the detection of *Taylorella equigenitalis*, the organism that causes Contagious Equine Metritis (CEM). The goal "is to demonstrate that the overall prevalence in the United States is very low (1 per 1,000 or less), if it exists at all. Results of the testing could be used to increase national and international confidence in the conclusion of the current outbreak response, and to potentially mitigate interstate and international reactions related to equine commerce should there be any future detection of *T. equigenitalis*."

New York will be testing approximately 100 stallions in the state. Enrollment of the stallions in this surveillance testing will be **voluntary**. Collection of the samples must be done by an accredited equine veterinarian. USDA-APHIS-VS will pay for the costs of the laboratory diagnostic testing as well as shipment of samples to a CEM approved laboratory but stallion owners are required to pay the accredited equine practitioner for sample collection costs.

The USDA-APHIS-VS surveillance study guidelines have been finalized. The guidelines explain which criteria would qualify stallions for testing, as well as testing costs and laboratory information. For more information about these guidelines and this surveillance study, please call Dr. Todd Johnson at the local USDA-APHIS-VS office at 518-218-7540.

The Animal Health Diagnostic Center at Cornell University is one of the CEM certified laboratories in the United States. If you need assistance on sampling, transport media or **general** information on CEM, contact the Diagnostic Laboratory at 607-253-3900.



New Tests for Johne's Disease Offered at AHDC

By Dr. Joy Bennett, DAI

The Animal Health Diagnostic Center (AHDC) at Cornell University, College of Veterinary Medicine, is now offering a direct fecal PCR (polymerase chain reaction) test for Johne's disease in cattle. This test has several advantages including the ability to reliably detect heavy and moderate shedders, quick turn-around time for results, and reasonable cost. These advantages have made this test a very popular option for herds seeking to control Johne's disease through management and testing. Submission of samples for direct fecal PCR testing requires scheduling with the lab. Please contact Monica Carey at the Johne's lab (607-266-8965) at least 4 weeks prior to submission. Advanced scheduling is not required for individual clinical suspect samples. This test is not currently approved for environmental samples, non-bovine species, or non-fecal specimens.

In addition, the AHDC is now offering a milk ELISA test for Johne's disease. This test is a commercial kit licensed to test milk for antibodies against *Mycobacterium avium* subsp. *paratuberculosis* (MAP). In accordance with the kit manufacturer's recommendations, results of this test are reported as POSITIVE or NEGATIVE with no quantitative qualifier attached. Individual cow milk samples may be submitted directly to the AHDC or through DairyOne. It is recommended that milk ELISA test results be interpreted at the herd level, rather than at the individual animal level.

For information on test selection, test performance and sampling procedures, please refer to Appendix G – Johne's Disease Program: Sampling and Testing Options, which can be found on the AHDC website at: <http://diaglab.vet.cornell.edu/test/feeman/AppendG.pdf>. Current Johne's test fees and comprehensive fact sheets on the direct fecal PCR test and the milk ELISA test are also available on the AHDC website.



Anaplasmosis

By Dr. Jim Gray, DAI

The policy concerning the handling of Anaplasmosis positive cattle has been revised as of November 2009 based on our experience with a shipment of unqualified cattle from Texas earlier in 2009. The cattle were dispersed to numerous locations after being sold through a New York livestock market. Based on significant testing of the imported and contact cattle, it was determined that our current policy could be safely modified as follows:

- Quarantine only the positive animals on the farm, not the entire farm
- Retest the positive animals 2 to 3 weeks after the initial test unless the animal is moved immediately to slaughter
- Release the quarantine upon a negative test result or proof of animal sent to slaughter
- Treat with long acting tetracycline (LA-200) to eliminate the carrier state
- Give 4 treatments at 3 day intervals at a dose of 20mg/kg (5 cc per 100 pounds) with no more than 10 cc's per IM injection site
- Retest treated animals as early as 30 days after the last treatment
- Consider meat and milk withholding times
- No indemnity will be paid for positive animals and NYSDAM cannot order their destruction
- Asymptomatic test positive cattle do not *need* to be kept separate from the herd
- Separate and treat or cull symptomatic animals
- Establish a herd control program with the herd veterinarian
- Advise owner to use individual needles on all animals and to properly disinfect equipment used between animals
- Notify DAI if quarantined animals are moved to slaughter or preapproved NY premises
- Out of state movements require preapproval from the receiving state



Going Green at the Division of Animal Industry

The NYS Department of Agriculture and Markets is going green! The department wants to do its part to help the environment by reducing its carbon footprint. So in an effort to reduce its carbon footprint, the divisions within the department have been asked to reduce printing and mailing of documents that can be found online. The Division of Animal Industry (DAI) continues to work to make their website more user friendly and accurate (www.agmkt.state.ny.us/AI/AIHome.html). Many DAI laws are now online. There is also a new webpage for the importation and exportation of animals in/out of New York. Lastly, the accredited veterinary newsletter will only be offered in an electronic format. Veterinarians who supply their email address to the Division of Animal Industry will receive this newsletter as well as other animal health alerts by email. If you or a colleague are not receiving the electronic newsletter or electronic animal health alerts and would like to, please contact our office at (518) 457-3502.



2010 Fair Information

By Dr. Jim Gray, DAI

There are no significant changes in the animal health requirements for the 2010 fair season. The BVD test requirement for cattle, llamas and alpacas remains in place. Please remember to list the BVD test date on the certificate of veterinary inspection (CVI). We strongly recommend that you use the latest version of the "Exhibition CVI", or AI-61, dated January '08. They are available from our office. Please call Gerard at 518-457-3502 to order those forms.

We have also recommended to the fair officials that swine and poultry be housed separately to minimize the chance of spread of avian influenza. We have also issued an H1N1 advisory. The advisory recommends that signs be posted advising people with symptoms of influenza to avoid animal areas to minimize the spread of H1N1 to animals, particularly swine and poultry. The Animal Health Requirement Booklet is posted on the DAI webpage (www.agmkt.state.ny.us/AI/AIHome.html) along with information the H1N1 advisory.

From the Desk of the Import/Export Clerk— Cattle Ear Tags & Movement to Canada

There has been some concern about recording the proper identification on international health certificates for cattle going to Canada. With the increasing types of identification that are being used by producers, it is important to be aware of which RFID tags will be allowable as official ID for entry into Canada. When in doubt, it is best to call the local USDA-APHIS-VS NY office at (518) 218-7540 well in advance of export to be sure that the given identification is properly recorded and allowable by Canadian officials at the border crossings.

The link below is for the most current list of USDA approved "840" ID devices

<http://www.aphis.usda.gov/newsroom/content/2008/10/naisvtag.shtml>

Cattle destined for Canada must meet these requirements:

<http://www.inspection.gc.ca/english/anima/heasan/pol/ie-2007-4e.shtml>

Remember that Canada (CFIA) wants US exporters from NY to use metal ("21") NY tags or "840" tags **with RFID**. There are a few 840 tags that lack RFID; these are inadequate for cattle exports to Canada.





CEM and Biosecurity Measures for the Equine Practitioner

By Dr. Linda Mittel, AHDC

Breeding season has begun for many breeds or is imminent for others even though the NY weather has been cold and snowy. For many equine "repro" veterinarians, it is a good time to make plans and organize your breeding protocols for the coming breeding season.

CEM (contagious equine metritis), a foreign animal disease, was detected in the US in 2008. After the USDA-APHIS tracebacks, it now is apparent that CEM has been in the US since 2004. New information indicates that CEM can be transmitted by fomites (phantoms, AVs, grooming tools, etc.), in transported cool semen, placental and fetal fluids, gelding's sheaths, tease horses and by the act of natural breeding. This information dictates that strict biosecurity measures should be part of every equine veterinarian's protocols.

The newly discovered modes of transmission will necessitate that biosecurity changes be made in certain routine procedures and **all** reproductive procedures. These measures should include the use of exam gloves and PPE (personal protective equipment) such as disposable or washable coveralls when handling any stallion, gelding or mare for reproductive/genital procedures. Common procedures such as castrations, sheath cleaning, foaling, tail wrapping, and foal handling require similar biosecurity measures. After each procedure PPE should be changed. This will require having multiple sets of PPE in your vehicle or in your clinic. Examining multiple mares at one farm can be difficult. Ultrasounds probes and cords should be cleaned and disinfected between each mare with a 2% chlorhexadine solution or covered with a rectal sleeve and changed between mares. The veterinarian needs to be cognizant of biosecurity and the possibility of transmitting CEM.

Mare infertility is one part of CEM infection, but mares do not remain infertile in all cases. Mares are known to transmit CEM to their foal at birth. Geldings have been found to also to carry the CEM organism (probably from birth) unless there was a history of breeding or other exposures prior to castration.

Clients will look to their veterinarians for assistance and guidance on how to determine if their stallion is free from CEM and how to market this health status to mare owners. There are no specific testing protocols to follow other than the USDA Import Regulations. They are very expensive and time consuming for the average horse owner. The AAEP Infectious Disease Committee is working on testing recommendations but they will not be available for this breeding season.

Most domestic stallion owners cannot follow the import testing protocols even though they want to test their stallions. Vets can recommend that the stallion be cultured before, during and after the breeding season as surveillance. This may be an effective surveillance tool especially if the stallion breeds any mares naturally or stands with other stallions. The APHIS CEM Coordination Group recommends three sites on the external genitalia to be cultured. The three sites are the urethral sinus, fossa glandis and distal urethra.

Semen cultures are not the best sample to detect the CEM organism because *Taylorella equigenitalis*, the bacterial agent of CEM, is a commensal that is found on the surface of the skin of the sheath and penis. It is usually not found in the semen but collected semen can be contaminated. Semen should be cultured however as part of the pre-breeding work up along with the aforementioned samples. Stallions can be cultured for both CEM and other aerobic pathogens at the same time. Duplicate swabs are necessary for cultures. If the stallion has been on antibiotics within 7 days, or has had topical antibiotics on the penis or prepuce within 21 days of culture, CEM cultures should not be taken.

(Continued on Page 6)



(Continued from Page 5 CEM and Biosecurity)

Specific transport media (Amies with charcoal) is required for CEM cultures. Samples must arrive at the lab and be set on culture media by 48 hours post-sampling. Only CEM approved laboratories should perform the CEM cultures and prior arrangements should be made so as to ensure that the laboratory has space. CEM cultures are finalized after 7 days, due to the slow growth of *Taylorella equigenitalis*. The Animal Health Diagnostic Center at Cornell is approved to do this testing.

Some practical considerations for your clients include:

- Using gloves in **all** reproductive procedures---Tail wrapping, washing mares and stallions, handling post foaling placental and fetus fluids
- Cleaning stalls thoroughly post foaling
- Providing breeding animals with their own grooming tools including sponges and buckets
- Using bucket liners when washing stallions and mares for either breeding or non breeding activities
- Recording the date that the stallion was cultured negative for CEM as well as approved laboratory name on shipped, cooled, semen paperwork
- Using the CF (complement fixation) blood tests to test mares post breeding
- Understanding that infected mares will seroconvert by 21 days post-breeding but do not persist
- Understanding that CF blood tests taken 45 days or later post breeding are often false negatives and if mares have previously been infected the current CF test may also be false negative
- Culturing mares with vaginal discharge post-breeding or a history of infertility

These considerations are meant to guide your recommendations for each farm, depending on the farm's activities and concerns. If you need additional assistance in designing testing protocols, assistance with CEM information or breeding surveillance plans, call the AHDC at Cornell at 607-253-3900. Request to speak to one of veterinary support service members. Sampling supplies are available through the shipping department at the AHDC at 607-253-3935.

PPD Tuberculin SHORTAGE

Due to a nationwide shortage of PPD tuberculin, the NYS Dept of Ag & Markets - Division of Animal Industry has a limited supply of tuberculin in our office. We are only able to stock small quantities. We have a few 10ml vials (100 tests), 5ml vials (40 tests) and 1ml vials (5 tests). If you have TB testing already scheduled and you are in need of PPD tuberculin, you need to call our office immediately at 518-457-3502. Further, if you foresee a need for tuberculin in the future (dispersal sale, interstate movement), please notify our office as soon as possible. We will need to know the approximate number of animals to be tested as we may only order as much as necessary. We will let you know when the national supply is no longer in shortage. Thank you for your help in this matter.

The Division of Animal Industry has a new
IMPORT/EXPORT webpage to help assist in moving
animals in and out of NY!

[http://www.agmkt.state.ny.us/ai/
import_export.html](http://www.agmkt.state.ny.us/ai/import_export.html)



Rabies and Vesicles in a Calf

By DAI Field Staff

On a late Friday afternoon in October 2009, the DAI received a call about a 50 cow dairy farm in the Finger Lakes area that had several, unexplained calf deaths in a short time frame. The first death was a 4 month old bull calf who had increased drooling, vocalization and dysphagia. The vet treated this calf early on for a possible necrotic pharyngitis. In time the calf became increasingly aggressive; running into gates and walls, and biting on pipes. Due to the deterioration of health, the calf was humanely euthanized by the owner on 10/2.

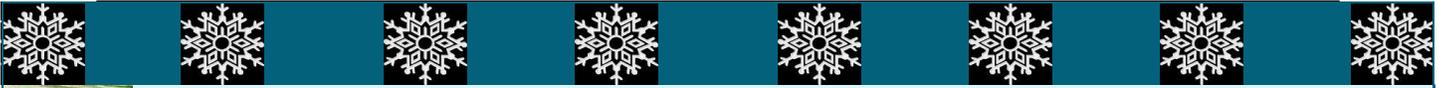
The second calf that died, a 6 month old heifer, had different symptoms. The heifer had progressive lameness starting in the rear limbs. As time went on, the heifer was knuckling at the fetlock and was eventually paralyzed in the rear limbs. She refused to eat despite intensive care including stomach tubing by the owner several times a day. She was found dead on 10/16.

The third and final calf that died started acting sick on 10/18. She started to drool and was unable to eat and drink easily. She also started to run into walls and gates. According to the owner, the calf acted 'rambunctious.' The herd vet who unsuccessfully treated her for possible Clostridial disease or Listeriosis also was suspicious of Rabies. On 10/23 the vet convinced the owner to have the ailing heifer calf submitted to Cornell University's Animal Health Diagnostic Center for necropsy.

The necropsy of the calf led to an unexpected turn of events. Due to the fact that Rabies was on the vet's differential diagnoses list, the calf's brain was submitted to NYSDOH's Wadsworth Center for Rabies testing. A full necropsy was also performed. It revealed that there was some mild dehydration as well as typhlocolitis with intraluminal hemorrhage. All other systems seemed rather unremarkable upon gross exam. However when the mouth and feet were examined, the vets noticed some unexpected lesions. The calf's mouth had a few vesicles and evidence of ulceration on the gingiva. On the coronary bands of both hind limbs, there were ulcerated areas of skin. Due to the fact that oral vesicles and coronary band ulcers can be indicative of Foot and Mouth Disease or Vesicular Stomatitis Virus, a foreign animal disease investigation was initiated. An FADD (foreign animal disease diagnostician) obtained the appropriate samples for the investigation and sent the samples to the FADDL (Foreign Animal Disease Diagnostic Lab) on Plum Island, NY. In the meantime, a BVD test was performed at Cornell. The BVD test was negative.

A field vet was called once it was determined to be a foreign animal disease investigation despite the fact that Rabies was a strong suspicion. The field vet went out to the farm the same night of the necropsy (10/23). The owner and herd veterinarian were updated on the necropsy findings and unexpected turn of events. Upon further discussion between the veterinarians and owner, it was learned that a dead, juvenile skunk had been found in the heifer barn back in August 2009. All three calves that died had been housed in the same heifer barn when the skunk was found. There had been no new outside additions to the herd or history of foreign travel by farm workers. The involved parties discussed all of the different possible diagnoses and determined that Rabies disease could be a very real possibility on the farm. The owner was given precautions to follow in the vent that any other cases should develop. The local county health department was contacted that same night and chose to initiate post-exposure Rabies treatment. The Wadsworth Center reported a positive diagnosis for Rabies over the weekend. Final results from FADDL on Plum Island, NY, were negative for both Foot and Mouth Disease and Vesicular Stomatitis Virus.

Even though this story may seem like a straight forward case of Rabies, there are a few interesting points to remember. First, even if you think that you know the diagnosis, there can be unexpected twists, like FAD symptoms (vesicles, ulcers). Don't ignore these findings even if it doesn't agree with your initial suspicions. Call state or federal officials. Second, some diseases like Rabies can occur in multiple animals even though usually a single animal is affected. Third, animals who are neurologic should be treated as rabies suspects. Humans who must interact with these animals should report this to state or federal officials and consider this a public health threat and take the necessary precautions.



Donkey Deaths

By DAI Field Staff

Late in 2009 a private veterinary practitioner was called to a beef farm because several donkeys, sharing a pasture with the farm's beef herd, were found dead and dying. Signs were acute in nature and included profound lethargy and cardiovascular collapse. Five out of six animals died within a very short period of time (2 days). The sixth donkey (moribund) was euthanized just prior to necropsy. All necropsies were conducted at NYS College of Veterinary Medicine. A pre-weaned male remains alive and, to date, has not shown signs of illness.

Poisoning or a highly virulent infectious disease (including a foreign animal disease), were on the list of possible causes. Necropsy and histopathology seemed to indicate a non infectious etiology. Given the history, review of premises, conversations with the owner, clinical signs, cardiac histopathology and known sensitivity of equine species to ionophores, concern centered on the possibility of ionophore toxicity. The investigation then narrowed in on the pasture and other available feeds.

The donkeys had access to the same pasture (currently scant due to the season) for over 5 years. In addition to pasture, the beef herd and donkeys were offered two commercial beef cattle mineral mixes (free choice) and hay. One or more mineral formulations were available at pasture feeding stations. Samples were taken from all feeding stations and from unopened bags of the same products. Samples were analyzed by NYS Department of Agriculture's Food Laboratory, Cornell's toxicology lab and Food and Drug Administration's laboratory. All laboratories reported finding trace levels of ionophores (monensin sodium and/or lasalocid) in all of the mineral formulation samples that were tested. No ionophores were detected in stomach or cecal contents. Selenium levels were determined (FDA) from all samples and found to be consistent with product label specifications.

None of the mineral mix labels listed ionophores as a component. The levels of lasalocid (min= .1 ugm/gm-feeder & max= 0.8 ugm/gm-feeders) and monensin sodium (min= 0.1 ugm/gm & max= 1.2 ugm/gm-feeders), identified in the mineral mix samples, were well below levels believed to cause acute toxicosis². Studies to determine toxic levels in donkeys could not be found but it is generally believed they would be similar to those in horses.

Although history, clinical signs and histopathology all support ionophore poisoning, sample analysis does not prove ionophore toxicosis. It may be that the samples that were analyzed do not reflect the level of ionophores that the donkeys actually consumed (another bag may have had very high levels) or, there may have been some type of additive effect from having more than one ionophore present. It is known, that certain antibiotics can potentiate the toxic effects of ionophores in other species.^{3,4}

While a definitive cause for the death of the donkeys could not be determined, this case emphasizes the importance of following label directions when feeding livestock. Supplements and additives meant for one species may not be tolerated by another species. Also, manufacturing practices for a product meant for one species may present a risk to another species. For cattle feeds, there may be an occasion where a formulation containing ionophores moves through the mixing system prior to a formulation that does not contain ionophores, at the very least, it appears that may have happened in this case.

1. Ralsbeck MF. 1992. Feed-associated poisoning. In: Current Therapy in Equine Medicine. 3rd ed. NE Robinson ed. W.B. Saunders, Philadelphia Pa. pp. 366-377.

2. Technical Bulletin, 2002, Animal Health Division Alpha Pharma Inc. One Executive Drive Fort Lee, NJ 07024 USA

3. Journal of Veterinary Diagnostic Investigation, Vol 11, Issue 1, 79-86 Copyright © 1999 by American Association of Veterinary Laboratory Diagnosticians Toxicosis in cattle from concurrent feeding of monensin and dried distiller's grains contaminated with macrolide antibiotics: RJ Basaraba, FW Oehme, MW Vorhies, and GL Stokka Department of Diagnostic Medicine/Pathobiology, Kansas State University, Manhattan 66506

4. An Outbreak Investigation of Ionophore Toxicity in a Group of Jersey Heifers Annie Stokowski Advisor: Dr. Gillian Perkins Clinician: Dr. Monica Figueiredo Senior Seminar Paper Cornell University College of Veterinary Medicine, January 22, 2003

